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Indirect plant regeneration in aromatic rice (*Oryza sativa* L.) var. 'Kalijira' and 'Chinigura'

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ABSTRACT

Mature seeds of two traditional rice genotypes (Kalijira and Chinigura) were used for callus induction and plant regeneration on different concentrations and combinations of plant growth regulators cultured on MS (Murashige and Skoog) basal medium. Callus induction frequency was different between the cultivars, as well as among the 2,4-dichlorophenoxyacetic acid (2,4-D) levels tested. Both tested cultivars exhibited highest callus frequency at 2 mg l⁻¹ 2,4-D. The incorporation of benzylaminopurine (BAP) and kinetin (KIN) in the callus induction medium supplemented with 2 mg l⁻¹ 2,4-D did not significantly improve the callus induction frequency but required days of callus initiation were decreased compared to single use of 2,4-D. After two subcultures, at 21 days interval, embryogenic callus was placed on medium containing different concentration and combination of auxin and cytokinin. Treatment T₄ (0.5 mg l⁻¹ BAP and 0.1 mg l⁻¹ IBA) showed the highest shoot induction: 91.67% in Kalijira and 83.33% in Chinigura. Similarly, the highest range of shoot number was also observed in both genotypes when treated with 0.5 mg l⁻¹ BAP and 0.1 mg l⁻¹ IBA. Plant regeneration efficiency was further observed best when treated with 1 mg l⁻¹ 2,4-D along with 1 mg l⁻¹ 2,4-D along with 1 mg l⁻¹ BAP and 1 mg l⁻¹ IBA. Furthermore, the highest number of callus derived shoot per culture was achieved in 2 mg l⁻¹ 2,4-D along with 1 mg l⁻¹ BAP and 1 mg l⁻¹ IBA. Both rice genotypes are promising in terms of callus induction frequency and morphology, and regeneration ability of the embryogenic callus.

Key words: callus induction, plant regeneration, aromatic rice, shoots

IZVLEČEK

POSREDNA REGENERACIJA AROMATIČNEGA RIŽA (*Oryza sativa* L.), SORT 'KALIJIRA' IN 'CHINIGURA'

Zrela semena dveh tradicionalnih genotipov riža ('Kalijira' and 'Chinigura') so bila uporabljena za indukcijo kalusa in regeneracijo rastlin pri različnih koncentracijah in kombinacijah rastlinskih rastnih regulatorjev pri gojenju na osnovnem MS (Murashige and Skoog) mediju. Frekvenca indukcije kalusa je bila različna med sortama kot tudi glede na koncentracije 2,4-diklorfenoksi očetne kisline (2,4-D). Obe preiskušeni sorti sta imeli največjo frekvenco kalusa pri 2 mg l⁻¹ 2,4-D. Dodatek benzilaminopurina (BAP) in kinetina (KIN) v medij za indukcijo kalusa z dodatkom 2 mg l⁻¹ 2,4-D ni značilno izboljšal indukcije kalusa, vendar so se potrebni dnevi za začetek tvorbe kalusa zmanjšali v primerjavi s postopkom, ko smo uporabili samo 2,4-D. Po dveh predkulturah, v interval 21 dni, je bil embriogeni kalus prenešen na medij, ki je vseboval različno koncentracijo in kombinacijo aoksina in citokinina. Tretma T₄ (0.5 mg l⁻¹ BAP in 0.1 mg l⁻¹ IBA) je dal največjo indukcijo poganjkov: 91.67 % pri 'Kalijira' in 83.33 % pri 'Chinigura'. Podobno je nastalo največ poganjkov pri obeh sortah, kadar so jih tretirali z 0.5 mg l⁻¹ BAP in 0.1 mg l⁻¹ IBA. Nadalje je bila sposobnost regeneracije rastlin najboljša, če so jih tretirali z 1 mg l⁻¹ 2,4-D z dodatkom 1 mg l⁻¹ BAP in 1 mg l⁻¹ IBA. Največje število iz kalusa nastalih poganjkov na kulturo je bilo doseženo pri 2 mg l⁻¹ 2,4-D z dodatkom 1 mg l⁻¹ BAP in 1 mg l⁻¹ IBA. Oba genotipa riža sta obetavna v smislu morfologije in pogostosti indukcije kalusa kot tudi v regeneracijski sposobnosti embriogenega kalusa.

Ključne besede: indukcija kalusa, regeneracija rastlin, aromatični riž, poganjki

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1 INTRODUCTION

Global population is increasing very rapidly. Loss in crop production could lead to hunger and famine, especially in the developing countries. So it is time to tackle the challenges of a rapidly increasing population and stiffer global competition in the next millennium. Moreover these two challenges require better research to produce more and better quality food efficiently. The improvement can possibly be achieved by creating genetic variability. Rice (*Oryza sativa* L.) is the world most important food supplier cereal crop after wheat and maize (Ray, 1985). It provides half of total dietary carbohydrate, especially in Asian countries and it is suitable diet for more than three billion people, supplying 50-80% of their daily calorie intake (Khush, 2005). Thus a considerable improvement has been done through traditional rice breeding. Rice breeding has made significant progress towards higher yield, improved quality, greater disease resistance and other important characters of agricultural importance in the past and even in future, it will still play an important role. Due to its increasing importance in nutrition and economy, it is now felt that new varieties of rice, having good agronomic characters, should be evolved.

Kalijira and Chinigura are the most important aromatic rice varieties of Bangladesh and the rest of the world due to its attractive flavor, fine grain and good taste. Aroma and taste are caused by the chemical compound 2-acetyl-1-pyrroline (Ghareyazie *et al.*, 1997). This rice is generally used to prepare dishes such as *polau*, *biriani* and different types of cake which are served on special occasions. Aromatic rice receives premium price

and is profitable for the growers as well as the traders. Country can benefit by earning exchange by production and export of aromatic rice.

Most of the aromatic rice cultivars are traditional rice varieties which have tall stature, low yield, photoperiod-sensitivity, are susceptible to disease and pest and unresponsive to fertilizer. But due to the favorite flavor and some other dominant grain quality characteristics, they are the important resource for breeding and improving the aromatic rice cultivars for diverse demands of consumers in the world. Several laboratories have described regeneration of plants from various rice explants such as immature embryos, immature panicles (Ling *et al.*, 1983), young inflorescence (Chen *et al.*, 1985) and root (Abe and Futsuhara, 1985). Rashid *et al.* (2000) studied that rice seeds have more potential for callogenesis as compared to node or tip. Successful callus induction from rice seed has been reported by several researchers (Gonalz, 2000; Alam *et al.*, 2003; Shahsavari *et al.*, 2010). The use of mature seeds has the advantage, because they can be obtained at anytime throughout the year regardless of growing season (Alam, 1994). Despite the enormous importance of aromatics rice, knowledge on the *in vitro* propagation of these rice lines is still elusive.

Therefore, this study was aimed at evaluating two Bangladeshi aromatic rice genotypes (Kalijira and Chinigura) for callus induction and regeneration efficiency under different concentrations and combinations of growth regulators.

2 MATERIALS AND METHODS

2.1 Explant sterilization and culture establishment

Mature seeds of two genotypes of aromatic rice namely; Kalijira and Chinigura were dehusked and immersed in 70% ethanol for 3 min, after washing the explants were dipped in 0.1% HgCl₂ solution for 3 minutes. The seeds were then rinsed 5-6 times with sterile distilled water to remove HgCl₂ with vigorous agitation in the laminar air flow cabinet. After surface sterilization of seeds, they were kept on autoclaved filter paper on the

petridish. When the water removed from the seeds surface it was inoculated into the culture tubes with sterilized forceps. The seeds were then placed on callus induction media and kept in the dark at 26 ± 2°C. MS (Murashige and Skoog, 1962) basal medium was used for callus induction and plant regeneration. In this study, 30 mg l⁻¹ sugars was used and solidified with 0.8% agar. The pH of the medium was adjusted to 5.8.

2.2 Callus induction

Different concentrations of 2,4-D (1, 2, 3 and 4 mg l⁻¹) were added into the MS medium for callus induction. Subculture was performed twice at 21-day interval using the same medium. Combinations of auxin and cytokinin (T₁=2.0 mg l⁻¹ 2,4-D+0.25 mg l⁻¹ KIN, T₂=2.0 mg l⁻¹ 2,4-D+0.5 mg l⁻¹ KIN, T₃=2.0 mg l⁻¹ 2,4-D+1.0 mg l⁻¹ KIN, T₄=2.0 mg l⁻¹ 2,4-D+1.5 mg l⁻¹ KIN, T₅=2.0 mg l⁻¹ 2,4-D+0.25 mg l⁻¹ BAP, T₆=2.0 mg l⁻¹ 2,4-D+0.5 mg l⁻¹ BAP, T₇=2.0 mg l⁻¹ 2,4-D+1.0 mg l⁻¹ BAP, T₈=2.0 mg l⁻¹ 2,4-D+1.5 mg l⁻¹ BAP mg l⁻¹) were also used in MS media for callus induction.

2.3 Plant regeneration

Embryogenic calli produced on MS medium containing 2 mg l⁻¹ 2,4-D were cultured on different regeneration media for plantlet formation. MS basal media supplemented with different concentrations and combinations of cytokinin and auxins (T₁=0, T₂=0.1 mg l⁻¹ IBA, T₃=0.1 mg l⁻¹ BAP+0.1 mg l⁻¹ IBA, T₄=0.5 mg l⁻¹ BAP+0.1 mg l⁻¹ IBA, T₅=1 mg l⁻¹ BAP+0.5 mg l⁻¹ IBA, T₆=0.5 mg l⁻¹ BAP+0.1 mg l⁻¹ IAA, T₇=0.5 mg l⁻¹ BAP+0.5 mg l⁻¹ IBA, T₈=3 mg l⁻¹ KIN+0.5 mg l⁻¹ NAA, T₉=3 mg l⁻¹ KIN+0.5 mg l⁻¹ IAA mg l⁻¹) were prepared for plantlet regeneration. Regenerated shoots were then transferred to half MS media immediately under light (2000 lux) provided by 40W white cool fluorescence tubes. The cultures were maintained in a growth chamber at 24 + 18°C for a 16 h photoperiod under cool white fluorescent lamps (Phillips Bangladesh Ltd.) and the light intensity

was maintained at 28–34 mol/m/s. Visual observation of culture was made every week.

2.4 Data recording

The frequency of callus induction and plant regeneration (%) were measured using the following formulas (Zaidi *et al.*, 2006):

$$\text{Frequency of callus induction (\%)} = \frac{\text{no. of explants induced callus}}{\text{no. of cultured explants}} \times 100$$

$$\text{Frequency of shoot induction (\%)} = \frac{\text{no. of culture induced shoot}}{\text{no. of culture}} \times 100$$

$$\text{Frequency of root induction (\%)} = \frac{\text{no. of shoot induced root}}{\text{no. of culture induced shoot}} \times 100$$

2.5 Statistical analysis

The experiments were arranged in a split plot design with three replications. Each replication per treatment contained 12 seeds for callus induction and 4-6 embryogenic calli for plant regeneration. Data were analyzed using the two way-factorial analysis of variance (factorial ANOVA), with plant growth regulator concentration as one treatment and genotype as the other treatment. Data were analyzed as means ± SE. IRRISate 7.2 software was also used to do ANOVA and DMRT.

3 RESULTS

3.1 Effect of 2,4-D on callus induction

Different concentrations (1.0, 2.0, 3.0 and 4.0 mg l⁻¹) of 2,4-D were used for producing sufficient amount of embryonic callus from mature seeds in MS medium. The results are presented in Table 1. Results indicate that growth regulators played a major role in callus induction. The callus induction was occurred at 7-12th days after inoculation.

Result showed that MS medium supplemented with 2 mg l⁻¹ of 2,4-D was most effective in callus induction in both Kalijira (97.22%) and Chinigura (94.44%). This indicates that the use of 2,4-D with 2 mg l⁻¹ was enough for production of high amount of callus in rice. Lowest range of days for callus induction was observed in both Kalijira and Chinigura in higher (4.0 mg l⁻¹) concentration of 2,4-D. The color of all Kalijira callus was creamy yellowish and Chinigura was creamy white but both of those textures were friable.

Table 1: Effect of 2,4-D in MS media on quality and quantity of callus induction.

Concentration of 2,4-D mg l ⁻¹	Kalijira			Chinigura			Mean (varieties)
	Range	%	Degree with callus morphology	Range	%	Degree with callus morphology	
1	10-12	91.66±1.3 a	+++Py,C	13-15	86.11±2.1 a	+++CrW,C	88.89±2.7
2	7-10	97.22±0.8 a	+++Py,C	11-13	94.44±0.8 a	+++CrW,C	95.83±1.4
3	7-10	94.44±0.8 a	+++Py,C	11-13	91.66±1.4 a	+++CrW,C	93.21±1.4
4	8-11	88.79±0.8 a	+++Py,C	12-15	88.78±0.8 a	++CrW,C	88.78±0.0
Mean (treatments)		93.0±1.7			90.2±1.7		

+ Slight callus, ++ Moderate callus, +++ Massive callus, Py= Pale yellow, C=Creamy and CrW= Creamy white; concentrations with the same letter were not significantly different at 0.05 probability level using LSD.

3.2 Effect of 2,4-D in combination with KIN and BAP on induce callus

Although 2,4-D (auxin) gave the highest result of callus induction in rice, some worker have showed a good result in other cereal crops (e.g. wheat) using 2,4-D in combination with low concentration of cytokinin.

The effect of cytokinin (BAP and KIN) along with (2,4-D, 2.0 mg l⁻¹) on callus induction was also tested in MS medium (result shown in Table 2).

KIN was found more effective (95.8±1.4) than BAP (94.4±0.0) for high amount of callus formation. In addition, numerous callus (95.8±1.4) was observed when explants were treated with 2.0 mg l⁻¹ of 2,4-D was supplemented with 0.5 mg l⁻¹ of KIN. Similar result was also found when treated with 2.0 mg l⁻¹ of 2,4-D (95.83 %). But required days of callus initiation were decreased (5-7 days) by using cytokinins along with 2,4-D than single use of 2,4-D in all cases.

Table 2: Effect of different combinations of growth regulator on callus initiation and callus growth.

Concentrations and combinations	Kalijira			Chinigura			mean
	Range	%	Degree	Range	%	Degree	
T ₁	6-7	94.44±0.8 a	+++	7-8	88.89±0.8 a	+++	91.6±2.7
T ₂	5-6	97.22±0.8 a	+++	5-7	94.44±0.8 a	+++	95.8±1.4
T ₃	5-6	88.89±2.1 a	+++	5-6	86.11±0.8 a	+++	87.5±1.4
T ₄	4-6	69.44±2.1 b	++	4-6	69.44±0.8 b	++	69.4±0.0
T ₅	7-8	91.67±1.4 a	+++	7-8	88.89±2.1 a	+++	90.2±1.4
T ₆	6-7	91.67±1.4 a	+++	7-8	91.67±1.4 a	+++	91.6±0.0
T ₇	5-7	94.44±0.8 a	+++	6-7	94.44±0.8 a	+++	94.4±0.0
T ₈	5-7	86.11±2.1 a	+++	5-7	83.33±2.4 a	+++	84.7±1.4
mean		88.23±3.08			86.15±2.8		

+ Slight callus, ++ Moderate callus, +++ Massive callus and same letter were not significantly different at 0.05 probability level using LSD. In each treatment, 36 explants were used.

3.3 Plantlet regeneration

The results indicate that, among different concentrations and combinations, treatment T₄ (0.5 mg l⁻¹ BAP +0.1 mg l⁻¹ IBA) showed better performance (Kalijira 91.67±0.18 and Chinigura 83.33±0.22) to produce plantlet while treatment T₂ (0.1 mg l⁻¹ IBA) shows the lowest results (Kalijira 41.67% and Chinigura 41.33%). The range of

shoot number (Kalijira 4-8 and Chinigura 3-8) and mean performance (Kalijira 6.63±0.18 and Chinigura 6.30±0.05) was also better for T₄ treatment and the lowest for T₂ (0.1 mg l⁻¹ IBA). Overall, Kalijira was found to be more efficient in producing plantlets than that of Chinigura.

Another experiment has been performed to find out the effect of 2,4-D on plant regeneration. Calli obtain from different concentration of 2,4-D (i.e. 1.0, 2.0, 3.0 and 4.0 mg l⁻¹) were used for regeneration.

The result showed that the highest percentages of callus producing shoot (40%) were observed from

the callus obtained from low concentration (Table 4). In addition, highest number of shoots was observed on the callus derived from 2.0 mg l⁻¹ of 2,4-D treatment. However, callus induced from high concentration of 2, 4-D (3.0 mg l⁻¹ or more) was found to be inefficient for plantlet regeneration.

Table 3: Regeneration efficiency of from callus derived from mature seeds (calli were obtained from 2.0 mg l⁻¹ of 2,4-D)

Treatments	Kalijira				Chinigura			
	Shoot induction (%)	Number of shoot		(% of shoots induced root)	Shoot induction (%)	Number of shoots		(% of shoots induced root)
		Range	$\bar{X} \pm SE$			Range	$\bar{X} \pm SE$	
T ₁	0	-	-	-	0	-	-	-
T ₂	41.67±0.26 e	1-4	2.80±0.20 c	100	41.33±0.62 e	2-3	2.60±0.23 f	100
T ₃	50.00±0.58 d	3-5	3.83±0.17 d	100	50.00±0.44 d	3-5	3.67±0.09 d	100
T ₄	91.67±0.63 a	4-8	6.63±0.18 a	100	83.33±0.22 a	3-8	6.30±0.05 a	100
T ₅	66.67±0.66 b	3-7	4.12±0.24 bcd	100	58.33±0.33 c	3-6	4.00±0.29 c	100
T ₆	50.00±0.88 d	2-6	3.83±0.00 d	100	50.00±0.58 d	2-5	3.00±0.29 e	100
T ₇	50.00±0.58 d	2-5	4.00±0.5 cd	100	41.67±0.55 e	2-5	3.20±0.11 e	100
T ₈	66.67±0.41 b	3-8	4.37±0.25 b	100	66.67±1.20 b	3-8	4.75±0.07 b	100
T ₉	58.33±1.01 c	3-7	4.28±0.42 bc	100	58.33±0.68 c	2-7	4.14±0.25 c	100
Mean (%)	59.37				56.25			

Treatments with the same letter were not significantly different at 0.05 probability level using LSD. In each treatment 12 explants were used.

Table 4: Plant regeneration efficiency of callus induced from different combinations of 2,4-D.

Calluses of different concentration of 2,4-D (mg l ⁻¹)	Concentration of BAP+IBA (mg l ⁻¹)	% of callus producing shoots	Average number of shoots per culture $\bar{X} \pm SE$	% of callus producing shoots	Average number of shoots per culture $\bar{X} \pm SE$	
			Kalijira	Chinigura		
1.0	0.5±0.1	40	4.5±0.29	40	4.0±0.40	
2.0	0.5±0.1	20	6.5±1.50	30	5.6±0.33	
3.0	0.5±0.1	20	3.5±0.50	20	3.5±0.50	
4.0	0.5±0.1	10	1.0±0.00	10	1.0±0.0	

4 DISCUSSION

Despite the great importance of aromatic rice, little information is available on the callus induction and plant regeneration method through *in vitro* culture. The present study investigated the effect of various growth regulators on callus induction and plant regeneration efficiency in two Bangladeshi Traditional Aromatic Rice var. Kalijira and Chinigura.

4.1 Callus induction

The results showed that MS medium supplemented with 2 mg l⁻¹ of 2,4-D was the most effective in callus induction for both cultivars Kalijira and Chinigura. This indicate that the use of 2,4-D with 2 mg l⁻¹ was adequate for production of high amount of callus in rice. This finding is in

agreement with previous report by Rashid *et al.* (2003), where they showed that Basmati 370, Basmati 385 and KS 282 produced high amount of callus cultured on MS medium supplemented with 2.0 mg l⁻¹ 2,4-D. Sikder *et al.* (2006) also reported that 2.0 mg l⁻¹ 2,4-D is better for Chinigura callus induction. Similar results were also found in Thai aromatic rice KDML105 (Summart *et al.*, 2008), ASD 16, ADT 43, Basmati 370, Pusa Basmati and Pokkali (Revathi and Pillai, 2011; Libin *et al.*, 2012; Islam *et al.*, 2005). Our results further revealed that the use of 2,4-D with cytokinin could be helpful for high and early production of callus. Similar observations were also reported in rice (Alam, 1994; Khondokar, 1999). Taken together, the findings from this study will be very useful for producing high frequency callus induction that is the prime step for crop improvement or rapid propagation through biotechnological approaches.

4.2 Plant regeneration

Among different concentrations and combinations, treatment T₄ (0.5 mg l⁻¹ BAP +0.1 mg l⁻¹ IBA) showed better performance to produce plantlets. The range of shoot number and mean performance

was also found to be better for T₄ treatment. Similar results were reported on indica (Khanna and Raina, 1998) and Japonica rice cultivars (Lee *et al.*, 2002). However, Sripichitt and Cheewasestatham (1994) reported that MS agar medium supplemented with 1 mg l⁻¹ indol-3-acetic acid (IAA) and 4 mg l⁻¹ benzyladenine (BA) induced highest percentage of calli forming shoots. Thadavong *et al.* (2002), Rashid *et al.* (2003), Sikder *et al.* (2006), Jubair *et al.* (2008) and Libin *et al.* (2012) also showed similar results. In our study, high concentrations of 2,4-D (2.0 mg l⁻¹ or more) were found to be suitable for callus induction but these calli were not efficient for plant regeneration. In this study, results also showed that Kalijira is more efficient than Chinigura for producing plantlet from callus.

Our findings provide a simple *in vitro* protocol for generating high frequency callus formation and its subsequent regeneration for aromatic rice. These findings can also be manipulated for disease and pest resistant variety, stress and salt tolerance variety through tissue culture and gene transfer techniques.

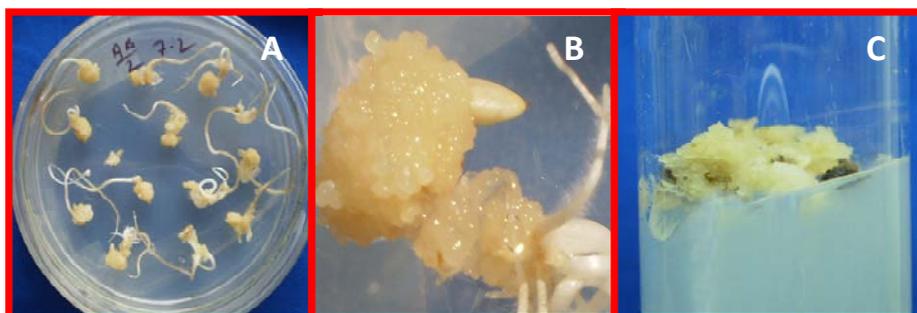


Figure 1: (A) Induction of callus from mature seeds in MS+2.0 mg l⁻¹ of 2,4-D. (B) Highlight a single seed derived callus. (C) Proliferation of callus.

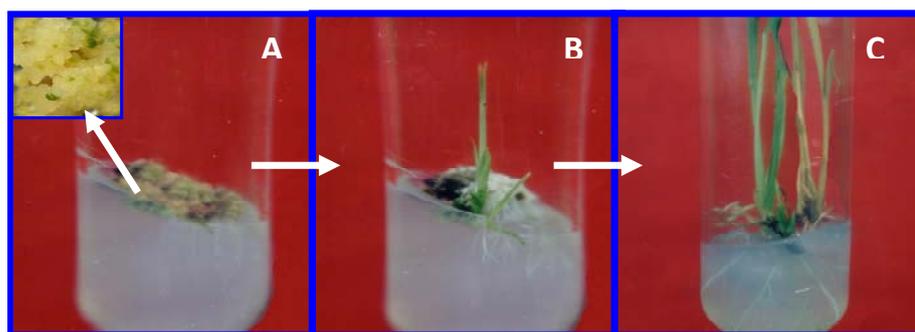


Figure 2: (A) Callus showing green spot on regeneration medium (MS +1.5 mg^l⁻¹ BAP +0.1 mg^l⁻¹ IBA). (B) Shoot formation from embryonic callus (MS+1.5 mg^l⁻¹ BAP+ 0.1 mg^l⁻¹ IBA). (C) Root proliferation of shoots in 1/2 MS without any growth regulators.

5 REFERENCES

- Abe, T.; Futsuhara, Y. (1985). Genotypic variability for callus formation and plant regeneration in rice (*Oryza sativa* L.). *Theoretical and Applied Genetics*, 72: 3-10.
- Alam M.F. (1994): Protoplast culture and transformation in rice (*Oryza sativa* L.). Ph.D. (Genetics) Thesis. Faculty of the Graduate School University of the Philippines, Lose Banos, Philippines.
- Alam, M. F.; Khatun, S. M.; Khandakar, I. A.; Khalekuzzaman, M.; Shohael, A. M. and Parvez, S. (2003). Genetic parameter on callus induction and plant regeneration using three explants in four rice cultivars of Bangladesh. *Bangladesh Journal of Genetics and Biotechnology*, 4(1&2): 59-62.
- Chen, T. H.; Lam, L. and Chen, S. C. (1985). Somatic embryogenesis and plant regeneration from cultured young inflorescence of *Oryza sativa* L. (rice). *Plant Cell, Tissue and Organ Culture*, 4: 51-54.
- Ghareyazie, B.; Alinia, F.; Menguito, C. A.; Rubia, L. G.; Palma, J. M. D.; Liwanag E. A.; Cohen, M. B.; Khush, G. S. and Bennett, J. (1997): Enhanced resistance to two stem borers in an aromatic rice containing a synthetic *cryIA(b)* gene. *Molecular Breeding*, 3: 401-414.
- Gonalz, M. C. (2000). Effects of different growth regulators on *in vitro* culture of rice cultivars. *Tropicales*, 21(1): 27-28.
- Islam, M. M.; Ahmed, M. and Mahaldar D. (2005). *In vitro* callus induction and plant regeneration in seed explants of rice (*Oryza sativa* L.). *Research Journal of Agriculture and Biological Sciences*, 1(1): 72-75.
- Jubair, T. A.; Salam, U.; Haque, N.; Akter, F.; Mukti, I. J.; Haque, A. K. M. F. and Ali M. R. (2008). Callus induction and regeneration of local rice (*Oryza sativa* L.) variety Topa. *Asian Journal of Plant Sciences*, 7(5):514-517.
- Khanna, H. K. and Raina S.K. (1998). Genotype x media culture interaction effects on regeneration response of three indica rice cultivars. *Plant Cell, Tissue and Organ Culture*, 52: 145-153.
- Khondokar, I. (1999). Callus induction and organogenesis of rice (*Oryza sativa* L.). M.Sc. Thesis, Department of Botany, University of Rajshahi, Bangladesh.
- Khush, G.S. (2005). What it will take to feed 5.0 billion rice consumers in 2030. *Plant Molecular Biology*, 59: 1-6.
- Lee, K., Jeon, H. and Kim, M. (2002.). Optimization of mature embryo-based *in vitro* culture system for high frequency somatic embryogenic callus induction and plant regeneration from Japonica rice cultivars. *Plant Cell, Tissue and Organ Culture*, 71: 237-244.
- Libin, A.; King, P. J. H.; Ong, K. H.; Chubo, J.K. and Sipe P. (2012). Callus induction and plant regeneration of Sarawak rice (*Oryza sativa* L.) variety Biris. *African Journal of Agricultural Research*, 7(30): 4260-4265.
- Ling, D. H.; Chen, W. F.; Chen, M. F. and Ma, Z. R. (1983). Direct development of plantlets from immature panicles of rice *in vitro*. *Plant Cell Reports*, 2: 172-174.
- Murashige, T. and Skoog F. (1962). A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Plant Physiology*, 15: 473-497.
- Rashid, H.; Abbasi, F.M. and Quraishi, A. (2003). Plant regeneration from seed derived callus of three

- varieties of basmati rice. *Plant Tissue Culture*, 13(1): 75-79.
- Rashid, H.; Toriyama, A.; Qurashi, K. And Malik, K. A. (2000). An improved method for shoot regeneration from calli of Indica rice. *Pakistan Journal of Biological Sciences*, 3(12): 2229-2231.
- Ray, J. K. (1985). Rice Research Institute in India. Indian Council of Agricultural Research, New Delhi, India. Introduction to Botany of the Rice Plant. 2nd Ed, p. 5.
- Revathi, S. and Pillai, M.A. (2011). *In vitro* callus induction in rice (*Oryza sativa* L.). *Research in Plant Biology*, 1(5): 13-15.
- Shahsavari, E.; Maheran, A. A.; Siti N. A. A. and Hanafi M. M. (2010). The effect of plant growth regulators on optimization of tissue culture system in Malaysian upland rice. *African Journal of Biotechnology*, 9(14): 2089-2094.
- Sikder, M. B. H.; Sen P. K.; Mamun, M. A.; Ali, M. R. and Rahman S. M. (2006). *In Vitro* Regeneration of Aromatic Rice (*Oryza sativa* L.) *International Journal of Agriculture & Biology*, 6: 759-762.
- Sripichitt, P. and Cheewasestatham P. (1994). Plant regeneration from embryo derived callus of aromatic rice (*Oryza sativa* L.) variety Khao Dawk Mali 105. *Kasetsart Journal Natural Science*, 28: 27-37.
- Summart, J.; Panichajakul, S.; Prathepha P. and Thanonkeo P. (2008). Callus induction and influence of culture condition and culture medium on growth of thai aromatic rice, Khao Dawk Mali 105, Cell Culture. *World Applied Sciences Journal*, 5(2): 246-251.
- Thadavong, S.; Sripichitt, P.; Wongyai, W. and Jompuk P. (2002). Callus induction and plant regeneration from mature embryos of glutinous rice (*Oryza sativa* L.) cultivar TDK1. *Kasetsart Journal: Natural Science*, 36: 334 – 344.
- Zaidi, M.A.; Narayanan, M.; Sardana, R.; Taga, I.; Postel, S.; Johns, R.; McNulty, M.; Mottiar, Y.; Mao, J.; Loit, E. and Altosaar, I. (2006). Optimizing tissue culture media for efficient transformation of different indica rice genotypes. *Agronomy Research*, 4: 563-575.